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# Supplementary Figure 1: The xCell development pipeline 



## Supplementary Figure 2: <br> Raw scores of the multiple signatures in the test samples

Sequencing-based testing samples
Adipocytes Astrocytes


B-cells
CD4+ memory T-cells



CD8+ T-cells



Chondrocytes



## Erythrocytes






HSC

ly Endothelial cells

Macrophages M1


Mast cells


1111111111111111111111111111111



Melanocytes




Monocytes



MSC
mv Endothelial cells





Osteoblast


Preadipocytes
$\left.1111111111111\right|_{111} 11111111111111$



Pericytes


Skeletal muscle


Smooth muscle




 N


OO

Tregs


Microarray-based testing samples

| adc | B-cells |
| :---: | :---: |
|  |  |
|  |  |
|  |  |
|  |  |



|  |  |
| :---: | :---: |
| 111111111111111111111111111111111111111111111 | 11111111111111111111111111111111111111111111 |
|  |  |



Class-switched memory B-cells


DC
Endothelial cells

|  |  |
| :---: | :---: |
|  |  |
|  |  |






Figure S2. Raw scores of the multiple signatures in the test samples. Each box plot shows the ssGSEA scores for a given set of a cell type's signatures (the signatures that correspond to the cell type presented in the title of the plot). The scores are shown on the 40 sequencing-based and 57 microarray-based left-out samples, which were not used for generating the signatures. In source of the sample is in parenthesis( $F$ FANTOM5, E - Encode, B - Blueprint, I - IRIS, N - Novershtern, H - HPCA). Box plots were colored to emphasize the cell type of interest (red), and its parental/descendants (green). Average of the scores is presented with a black circle. xCell uses the average of the multiple signatures in downstream analyses.

## Supplementary Figure 3:

## Simulated mixtures of pure cell types inferred by raw xCell scores

Blueprint - simulated mixture of $\mathbf{1 2}$ cell types


Blueprint - simulated mixture of 14 cell types


Blueprint - simulated mixture of 16 cell types


ENCODE - simulated mixture of $\mathbf{1 0}$ cell types


## FANTOM5 - simulated mixture of 11 cell types



FANTOM5 - simulated mixture of 15 cell types


IRIS - simulated mixture of $\mathbf{1 2}$ cell types

|  |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: |
|  |  |  |  |  |  |

Novershtern - simulated mixture of 14 cell types

|  |  |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  |  |  |  |  |  |  |

HPCA - simulated mixture of 14 cell types


HPCA - simulated mixture of 18 cell types


Figure S3. Simulated mixtures of pure cell types inferred by raw xCell scores. Each set of scatter plots represents 500 simulated mixtures of gene expression profiles of pure cell types chosen randomly from the training set. The simulation is performed as following: we chose a data source and a set of cell types that are found in this data source. For each of the 500 mixtures one of the several samples of the cell type is chosen by random, and its expression profile is then multiplied by a random fraction. The expression profile of the mixture is the sum of expression all cell types presented in the mixture. Each scatter plot shows the inferred raw score by xCell (average of all the cell type's corresponding signatures) in the x-axis compared to the underlying fraction of the cell type. Pearson correlation is presented. The $x$-axis is an enrichment score, and presents the full range of the scores, but in each cell type this range may be completely different. The plots show that in most cell types the raw enrichment scores are reliable in predict even small changes in the proportions of cell types. We notice however that in the sequencing-based data sources the association between the scores and the abundances are not linear.

## Supplementary Figure 4:

Transformation procedure of raw scores to linear scales




Figure S4. Transformation procedure of raw scores to linear scales. For each cell type we generated 126 simulated mixtures using 2 cell types - the corresponding cell type and one of two options - MPP (multipotent progenitor) cells or Endothelial cells. We used those cell types because they are found in all three sequencing-based methods (this procedure was performed for both seq-based and array-based samples, but show here only transformations of seq-based). Endothelial cells was used for the hematopoietic stem cells, and MPP for all other cell types. In these synthetic simulations the expression profile of the cell types that is used is the median gene expression across samples of the corresponding cell type. To fit a power function, we used only the simulations were the corresponding cell types abundances are between $0.8 \%$ to $25.6 \%$. We used this range because we are mostly interested in identifying cell types with low abundance, and above that the function exponential increase may interfere in a precise fitting. The raw scores were shifted to zero (by deducting the score of $0.08 \%$ ) and divided by 5000 . Each plot shows the fitted curve (in red, the black dots are the data points) and the curve after transforming it using the learned formula (if the cell type is available in multiple sources, the parameters are averaged). The learned formula is presented at the top.

## Supplementary Figure 5:

Cell types inferences in gene expression simulations using training samples
Blueprint simulation 1 (using training samples)


Comparison between methods


## Blueprint simulation 2 (using training samples)



Comparison between methods


## Blueprint simulation 3 (using training samples)



## ENCODE simulation (using training samples)





FANTOM5 simulation 1 (using training samples)


FANTOM5 simulation 2 (using training samples)


IRIS simulation (using training samples)


## Novershtern simulation (using training samples)



## HPCA simulation 1 (using training samples)



## HPCA simulation 2 (using training samples)



## Summary table of simulation based on training samples

\(\left.$$
\begin{array}{ccc|ccc}\hline & & \text { Diagonal } & & \text { Off-diagonal } & \begin{array}{c}\text { Adjusted } \\
\text { Adjusted }\end{array} \\
& \begin{array}{c}\text { Average R } \\
\text { (diagonal) } \\
\text { Non- }\end{array} & \begin{array}{c}\text { adjusted } \\
\text { adjusted } \\
\text { diagonal// } \\
\text { Non- } \\
\text { adjusted }\end{array} & \begin{array}{c}\text { Non- } \\
\text { associations } \\
\text { Numbero of }\end{array} & \begin{array}{c}\text { adjusted in } \\
\text { >0.25 }\end{array}
$$ <br>

associations\end{array}\right]\)|  | Blueprint 1 | 0.836 | 1.006 | 0.892 |
| :---: | :---: | :---: | :---: | :---: |
| Blueprint 2 | 0.797 | 1.019 | 0.899 | 4 |
| Blueprint 3 | 0.757 | 1.005 | 0.840 | 16 |
| ENCODE | 0.693 | 0.973 | 0.940 | 3 |

Figure S5. Cell types inferences in gene expression simulations using training samples. Each slide presents the results of xCell, published signatures and CIBERSORT, in predicting the underlying abundances of 500 simulated mixtures generated using the training samples. Each sample in a mixture set is generated by randomly choosing one of the multiple samples corresponding to each of the cell types including in the mixture. Top left: Pearson coefficients of xCell before applying the spillover compensation. Top right: Pearson coefficients of xCell after applying the spillover compensation. Bottom left: Pearson coefficients of published signatures, CIBERSORT. Bottom right: Comparison between all methods.

## Supplementary Figure 6:

Cell types inferences in gene expression simulations using testing samples
HPCA test simulation 1


Comparison between methods


## HPCA test simulation 2





## HPCA test simulation 3



Comparison between methods


## Novershtern test simulation 1



## Novershtern test simulation 2



## Novershtern test simulation 3



IRIS test simulation 1


## IRIS test simulation 2



IRIS test simulation 3


## Blueprint test simulation 1



## Blueprint test simulation 2



## Blueprint test simulation 3



ENCODE test simulation 1


ENCODE test simulation 2


ENCODE test simulation 3


Comparison between methods


## FANTOM5 test simulation 1



## Comparison between methods



## FANTOM5 test simulation 2



## FANTOM5 test simulation 3



Summary table of simulation based on training samples

|  | Diagonal |  |  | Off-diagonal |  |
| :---: | :---: | :---: | :---: | :---: | :---: |
|  |  |  |  |  | Adjusted <br> diagonal/ |
|  | Adjusted <br> Average R <br> (diagonal) | diagonal/ <br> Non-adjusted | Adjusted <br> diagonal/ <br> Non-adjusted | Number of <br> associations $>$ <br> Non-adjusted | in >0.25 <br> insociations |
| HPCA 1 | 0.849 | 1.015 | 0.921 | 5 | 0.997 |
| HPCA 2 | 0.872 | 1.049 | 0.926 | 6 | 0.879 |
| HPCA 3 | 0.904 | 1.023 | 0.822 | 3 | 0.686 |
| Novershtern 1 | 0.721 | 1.006 | 0.929 | 10 | 0.980 |
| Novershtern 2 | 0.614 | 0.944 | 0.926 | 10 | 0.900 |
| Novershtern 3 | 0.795 | 1.085 | 0.851 | 11 | 0.782 |
| IRIS 1 | 0.853 | 1.091 | 0.857 | 10 | 0.758 |
| IRIS 2 | 0.827 | 1.059 | 0.902 | 10 | 0.818 |
| IRIS 3 | 0.813 | 1.070 | 0.876 | 15 | 0.808 |
| Blueprint 1 | 0.874 | 0.994 | 0.921 | 3 | 0.704 |
| Blueprint 2 | 0.885 | 0.995 | 0.941 | 3 | 0.894 |
| Blueprint 3 | 0.906 | 1.013 | 0.944 | 4 | 0.846 |
| ENCODE 1 | 0.872 | 1.005 | 0.985 | 2 | 0.965 |
| ENCODE 2 | 0.854 | 1.015 | 0.945 | 1 | 0.978 |
| ENCODE 3 | 0.857 | 0.994 | 0.967 | 2 | 0.947 |
| FANTOM5 1 | 0.913 | 1.031 | 0.928 | 5 | 0.854 |
| FANTOM5 2 | 0.886 | 1.035 | 0.862 | 5 | 0.568 |
| FANTOM5 3 | 0.696 | 1.047 | 0.881 | 14 | 0.932 |
| Average | 0.860 | 1.014 | 0.930 |  | 0.854 |
|  |  |  |  |  |  |

Figure S6. Cell types inferences in gene expression simulations using testing samples. Each slide presents the results of xCell, published signatures and CIBERSORT, in predicting the underlying abundances of 250 simulated mixtures generated using the left-out testing samples, with 20\% noise. Top left: Pearson coefficients of xCell before applying the spillover compensation. Top right: Pearson coefficients of xCell after applying the spillover compensation. Bottom left: Pearson coefficients of published signatures, CIBERSORT. Bottom right: Comparison between all methods.

## Supplementary Figure 7:

Distributions of cell types' scores from random mixtures

## Sequencing-based distributions





## Array-based distributions




xCell scores
Figure S7. Distributions of cell types' scores from random mixtures. Beta distributions were learned from random mixtures excluding the cell type of interest in each of the 6 reference data sets (training samples only). The construction of the random mixture is described in the methods section. The distributions presented here, and are used for the statistical significance assessment, are combinations of the beta distributions FANTOM5, Blueprint and ENCODE for sequencing-based inputs, and IRIS, Novershtern and HPCA for array-based inputs.

## Supplementary Figure 8:

Dependencies between CD8+ T-cells and NK cells


Figure S8. Dependencies between CD8+ T-cells and NK cells. Gene signatures tend to not be reliable in differentiating between closely related cell types. Here we show an example of a dependency between CD8+ T-cells and NK cells using simulated mixtures of GSE60424 RNA-seq expression profiles (which were not part of the generation of the method). The top plot shows the correlations of 9 published signatures with CD4+ T-cells, CD8+ T-cells and NK cells. Only the Rooney signatures was able to reliably infer all three cell types. The bottom plots show a curve fitted to the scores of CD8+ T-cells in each of the methods. The red curves are association with CD8+ T-cells underlying abundances, while the blue curves are with the NK underlying abundances. The blue line is expected to be flat, while the red curve is expected to be linear. Both our method and CIBERSORT (Newman) perform well here. The Rooney signatures is not linearly associated with the abundance.

# Supplementary Figure 9: CD8+ T-cells scores vs. CD8A expression in cancer cell lines 



| Cell ID | Primary | Subtype | CD8A expression | Rank | $\begin{gathered} \text { CD8+ T-cells } \\ \text { scores } \\ \hline \end{gathered}$ | Rank |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Pfeiffer | H\&L | Diffuse Large B Cell Lymphoma | 7.968 | 1 | 0.47 | 16 |
| HPB-ALL | H\&L | Acute Lymphoblastic T Cell Leukaemia | 6.866 | 2 | 0.88 | 2 |
| SUP-T1 | H\&L | Acute Lymphoblastic T Cell Leukaemia | 5.723 | 3 | 0.74 | 5 |
| KE-37 | H\&L | Acute Lymphoblastic T Cell Leukaemia | 4.194 | 4 | 0.81 | 3 |
| ALL-SIL | H\&L | Acute Lymphoblastic T Cell Leukaemia | 4.058 | 5 | 0.67 | 7 |
| TALL-1 | H\&L | Acute Lymphoblastic T Cell Leukaemia | 3.884 | 6 | 0.69 | 6 |
| RI-1 | H\&L | B Cell Lymphoma Unspecified | 2.954 | 7 | 0.19 | 125 |
| CML-T1 | H\&L | Blast Phase Chronic Myeloid Leukaemia | 2.939 | 8 | 0.80 | 4 |
| PF-382 | H\&L | Acute Lymphoblastic T Cell Leukaemia | 2.67 | 9 | 0.49 | 14 |
| KCL-22 | H\&L | Blast Phase Chronic Myeloid Leukaemia | 2.318 | 10 | 0.27 | 45 |
| GSS | Stomach | Adenocarcinoma | 2.251 | 11 | 0.07 | 196 |
| NCI-H2227 | Lung | Small Cell Carcinoma | 2.217 | 12 | 0.06 | 232 |
| KASUMI-1 | H\&L | Acute Myeloid Leukaemia | 1.873 | 13 | 0.22 | 97 |
| HUH-6-clone5 | Liver | Hepatoblastoma | 1.859 | 14 | 0.03 | 419 |
| HARA | Lung | Squamous Cell Carcinoma | 1.819 | 15 | 0.04 | 296 |
| MOLT-16 | H\&L | Acute Lymphoblastic T Cell Leukaemia | 1.791 | 16 | 0.96 | 1 |
| DU-4475 | Breast | Ductal Carcinoma | 1.645 | 17 | 0.11 | 162 |
| MOLT-3 | Na | Na | 1.579 | 18 | 0.62 | 9 |
| DMS-79 | Lung | Small Cell Carcinoma | 1.56 | 19 | 0.02 | 567 |
| NCI-H522 | Lung | Non Small Cell Carcinoma | 1.559 | 20 | 0.08 | 190 |
| NCI-H1623 | Lung | Adenocarcinoma | 1.53 | 21 | 0.06 | 222 |
| CHP-126 | Autonomic Ganglia | Ns | 1.504 | 22 | 0.09 | 179 |
| MFE-280 | Endometrium | Adenocarcinoma | 1.498 | 23 | 0.02 | 591 |
| HuT 78 | H\&L | Mycosis Fungoides-Sezary Syndrome | 1.372 | 24 | 0.61 | 10 |
| NCI-H211 | Lung | Small Cell Carcinoma | 1.371 | 25 | 0.16 | 142 |

Figure S9. CD8+ T-cells scores vs. CD8A expression in cancer cell lines. Many methods rely solely on the expression of CD8A is a marker for CD8+ T-cells abundance. Here we exemplify the risk in doing so - using 929 CCLE RNA-seq expression profiles we calculated xCell scores and CD8A expression (top). We observed that CD8A is high in some non-T-cells originating tumors, and low in some T-cell leukemias. Looking at the top 25 cancer cell lines, only 7 of the 16 T-cell leukemia's are in the top of expression (bottom), while all of them are in top of $x$ Cell's CD8+ T-cells scores.

# Supplementary Figure 10: xCell scores in 37 TCGA \& TARGET cancer types <br> aDC 


Adipocytes

Astrocytes
 B-cells


CD4+ naive T-cells


CD4+ T-cells




 CD8+ Tcm


CD8+ Tem
 cDC
 Chondrocytes


 CMP
 DC
 Endothelial cells
 Eosinophils

Epithelial cells


Erythrocytes



Hepatocytes


HSC



Keratinocytes

ly Endothelial cells
 Macrophages



Macrophages M2


Megakaryocytes


Melanocytes


Memory B-cells


MEP

Mesangial cells






Neurons


Neutrophils




Osteoblast



Pericytes

Plasma cells

Platelets


Preadipocytes

pro B-cells


Sebocytes


Skeletal muscle




Th1 cells

Th2 cells



Figure S10. xCell scores in 37 TCGA \& TARGET cancer types. Box plots of cell types enrichment scores in 9,947 TCGA \& TARGET primary tumor samples across 37cancer types.

## Supplementary Figure 11: <br> Purity estimations using xCell scores



CPE purity


CPE purity

Figure S11. Purity estimations using xCell scores. We derived a microenvironment score as an. We correlated this score with the CPE purity measurements we previously generated using ESTIMATE, ABSOLUTE, LUMP and H\&E slides. In all cancer types, with uterine carcinosarcoma as the exception we observed high correlations between our new microenvironment score and CPE.

Supplementary Figure 12: t-SNE plots based on cell types scores

ACC


Lymphoid cell types ( $\mathrm{N}=21$ )



Figure S12. t-SNE plots based on cell types scores. Using the cell types inferences we generated t-SNE plots for 9,947 TCGA \& TARGET primary tumor samples across 37 cancer types. In each plot the analyses was performed using a subset of the 64 cell types. The cell types included in each set can be found in supplementary table 1. Each subset of cell types distinguish different cancer types from each other.

